

RESEARCH

Open Access



Species and genetic variability of sweet potato viruses in China

Yongjiang Wang^{1,2†}, Yanhong Qin^{1,2†}, Shuang Wang^{1,2}, Desheng Zhang^{1,2}, Yuting Tian^{1,2}, Fumei Zhao^{1,2}, Yingzhi Wang^{1,2}, Hui Lv^{1,2}, Qi Qiao^{1,2*} and Zhenchen Zhang^{1,2*}

Abstract

China is the world's largest producer of sweet potato (*Ipomoea batatas* (L.) Lam.). Considering that there are numerous sweet potato-producing regions in China and sweet potato is a vegetatively propagated crop, the genetic diversity of sweet potato viruses could be high in the country. However, studies on species and genetic variabilities of sweet potato viruses in China are limited, making it difficult to prevent and control viral diseases in this crop. During 2014–2019, sweet potato samples with viral disease-like symptoms were randomly collected from sweet potato fields in 25 provinces in China. Twenty-one virus species, including 12 DNA and 9 RNA viruses, were identified in the samples using next-generation sequencing, polymerase chain reaction and rolling-circle amplification methods. One novel sweepovirus species, *Sweet potato leaf curl Hubei virus* (SPLCHbV), was identified. Two species, *Sweet potato collusive virus* and *Tobacco mosaic virus*, were identified for the first time in sweet potato in China. Full-length or nearly full-length genomic sequences of 111 isolates belonging to 18 viral species were obtained. Genome sequence comparisons of potyvirus isolates obtained in this study indicate that the genome of sweet potato virus 2 is highly conserved, whereas the other four potyviruses, sweet potato feathery mottle virus, sweet potato virus G, sweet potato latent virus and sweet potato virus C, exhibited a high genetic variability. The similarities among the 40 sweepovirus genomic sequences obtained from eight sweepovirus species are 67.0–99.8%. The eight sweepoviruses include 14 strains, of which 4 novel strains were identified from SPLCHbV and 1 from sweet potato leaf curl Guangxi virus. Five sweet potato chlorotic stunt virus (SPCSV) isolates obtained belong to the WA strain, and the genome sequences of SPCSV are highly conserved. Together, this study for the first time comprehensively reports the variability of sweet potato viruses in China.

Keywords: Sweet potato virus, Species, Genetic diversity, China

Background

Sweet potato, an important food crop worldwide, ranks third in production and fifth in calorific contribution to human diet among all crops globally (de Albuquerque et al. 2019). China is the world's largest producer of sweet potato, with a plantation area of 2.37×10^6 ha and yield of 5.20×10^7 tons in 2019 (FAOSTAT 2019).

Sweet potato, a vegetatively propagated crop, can be infected by viruses during propagation, resulting in yield and quality reduction (Clark et al. 2012). Viral diseases severely affect sweet potato production, particularly sweet potato virus disease (SPVD) caused by co-infection with sweet potato feathery mottle virus (SPFMV) and sweet potato chlorotic stunt virus (SPCSV). Sweet potatoes with SPVD exhibit severe symptoms including stunting, leaf distortion, vein clearing and chlorosis, leading to an 80–90% decrease in production (Clark et al. 2012).

*Correspondence: qiaoq2005@126.com; zhangzhenchen@126.com

†Yongjiang Wang and Yanhong Qin have contributed equally to this work

¹ Institute of Plant Protection, Henan Academy of Agricultural Sciences, Zhengzhou, Henan, China

Full list of author information is available at the end of the article



© The Author(s) 2021. **Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit <http://creativecommons.org/licenses/by/4.0/>.

At least 30 viruses, including DNA and RNA viruses, have been found to infect sweet potatoes worldwide (Clark et al. 2012). Among them, at least 20 have been reported in China. The RNA viruses reported in China include SPFMV, sweet potato virus C (SPVC), sweet potato latent virus (SPLV), sweet potato virus G (SPVG) and sweet potato virus 2 (SPV2) of the genus *Potyvirus* (Colinet and Kummert 1993; Colinet et al. 1994, 1997, 1998; Ateka et al. 2007); SPCSV of the genus *Crinivirus* (Qiao et al. 2011); and sweet potato chlorotic fleck virus (SPCFV) of the genus *Carlavirus* (Aritua et al. 2009). Conversely, the DNA viruses reported in China include sweet potato leaf curl virus (SPLCV), sweet potato leaf curl China virus (SPLCCNV), sweet potato leaf curl Georgia virus (SPLCGoV), sweet potato leaf curl Canary virus (SPLCCV), sweet potato leaf curl Henan virus (SPLCHnV), sweet potato leaf curl Guangxi virus (SPLCGV), sweet potato leaf curl Sichuan virus 1 (SPLCSiV1), sweet potato leaf curl Sichuan virus 2 (SPLCSiV2) and sweet potato leaf curl Shandong virus (SPLCSdV), which all belong to the genus *Begomovirus* (Luan et al. 2006, 2007; Liu et al. 2013, 2014, 2017; Qin et al. 2013a). Begomoviruses include the following four groups: Old and New World viruses, “sweepoviruses” and “legumoviruses” (Zerbini et al. 2017). Sweepoviruses are phylogenetically distinct from Old and New World begomoviruses, and they infect plants of the *Convolvulaceae* family, including sweet potato (Hassan et al. 2016; Zerbini et al. 2017). Other DNA viruses infecting sweet potato in China include sweet potato badnavirus A (SPBV-A) and sweet potato badnavirus B (SPBV-B) of the genus *Badnavirus* (Li et al. 2018; Qin et al. 2016), and sweet potato symptomless virus 1 (SPSMV-1) of the genus *Mastrevirus* (Wang et al. 2015).

In China, sweet potato is produced in the northern, Yangtze River and southern regions, spanning tropical, subtropical, warm temperate and temperate zones and covering mountainous areas, hills, plains and coastal areas, and presents complex ecotypes. Accordingly, sweet potato viruses could be highly diverse in China. Practically, sweet potato viral disease control and prevention is a big challenge owing to a lack of comprehensive research on the casual viruses. Advances in molecular detection techniques of plant viruses, particularly next-generation sequencing (NGS), have provided critical tools for studying viral diversity (Kreuze et al. 2009). More than 100 novel DNA and RNA plant virus species have been discovered via NGS in recent years (Gu et al. 2014; Hadidi et al. 2016). The aim of the present study was to investigate the sweet potato virus species and their genetic variability using molecular detection techniques. Here, 21 virus species were identified in sweet potato samples in China, and genome variability of the major viruses was

analyzed. The results provide a basis for future research on the control and management of sweet potato viral diseases in China.

Results

Sweet potato virus species and their distribution in China

In this study, 21 virus species were identified in sweet potato samples using the NGS, PCR, RT-PCR and RCA methods (Table 1 and Additional file 1: Table S1). There are 12 DNA and 9 RNA viruses, belonging to nine genera and including one novel sweepovirus species, *Sweet potato leaf curl Hubei virus* (SPLCHbV). SPLCHbV was approved and ratified recently by the International Committee on Taxonomy of Viruses (ICTV), and has been listed in the ICTV website. In addition, to the best of our knowledge, here we identified tobacco mosaic virus (TMV) and sweet potato collusive virus (SPCV) in sweet potatoes for the first time in China. In our investigation, SPFMV was the most common virus infecting sweet potato and was detected in 23 provinces, followed by SPVG, SPLCV and SPCSV, which were detected in 22, 18 and 18 provinces, respectively (Table 1 and Additional file 1: Table S1).

Genetic diversity of potyviruses

Full-length or nearly full-length genomic sequences of 14 isolates of SPFMV were obtained from 9 provinces, 18 isolates of SPVG were obtained from 8 provinces, 10 isolates of SPLV were obtained from 4 provinces, 3 isolates of SPV2 were obtained from 1 province, and 11 isolates of SPVC were obtained from 5 provinces (Additional file 1: Table S2).

The phylogenetic analysis revealed that the 14 SPFMV isolates obtained in this study were classified into three major phytogroups A, B and C based on their polyprotein nucleotide (nt) sequences (Fig. 1 and Additional file 2: Figure S1). Pairwise comparisons of the polyprotein gene sequences of these 14 isolates revealed their nt and deduced amino acid (aa) sequence identities in the range of 86.0–100% and 92.2–99.9%, respectively. Nine out of the 14 isolates, namely, O-gui-17-5, O-ji-17-49, O-lu-17-8, O-qian-17-1, O-shaan-17-5, O-su-17-10, O-wan-17-1, O-wan-17-3 and O-yue-17-9, and reference isolates of the O-II minor phytogroup (strain O), including O-Ch1 (KY296450), O-10-O (AB439206) and O-Ordinary (AB465608), were grouped into one clade, indicating that these nine isolates belong to the O-II minor phytogroup. The nt and aa sequence identities among the O-II minor phytogroup isolates are 93.6–99.6% and 96.0–99.6%, respectively. The two isolates, RC-31-2 and RC-33-1, were clustered together with the B phytogroup (strain RC), RC-Ch1 and RC-S, suggesting that they belong to the B phytogroup. The

Table 1 Sweet potato virus species and their distribution in China

Species	Genus	GenBank no. of representative sequences	Distribution (provinces)	Methods
Sweet potato feathery mottle virus	Potyvirus	MK778785, MK778794, MK778796	Anhui, Beijing, Chongqing, Fujian, Gansu, Guangdong, Guangxi, Guizhou, Hainan, Henan, Hebei, Hubei, Hunan, Jiangsu, Jiangxi, Liaoning, Ningxia, Shandong, Shaanxi, Shanxi, Sichuan, Yunnan and Zhejiang	NGS, PCR
Sweet potato virus C	Potyvirus	MK778817	Chongqing, Fujian, Guangdong, Guangxi, Guizhou, Hainan, Hebei, Hubei, Jiangsu, Jiangxi, Shandong and Sichuan	NGS, PCR
Sweet potato virus G	Potyvirus	MK778826, MK778834, MK778835, MK778837, MK778838	Anhui, Beijing, Chongqing, Fujian, Guangxi, Gansu, Guangdong, Jiangxi, Guizhou, Hainan, Hebei, Henan, Hunan, Hubei, Jiangsu, Liaoning, Ningxia, Shaanxi, Shandong, Sichuan, Yunnan and Zhejiang	NGS, PCR
Sweet potato latent virus	Potyvirus	MK778808	Anhui, Chongqing, Guangdong, Hebei, Henan, Hubei, Hunan, Jiangsu, Jiangxi, Liaoning, Shaanxi, Shanxi, Shandong and Sichuan	NGS, PCR
Sweet potato virus 2	Potyvirus	MK778812	Anhui, Chongqing, Fujian, Hebei, Henan, Hunan, Jiangsu, Jiangxi, Liaoning, Shaanxi, Shanxi, Sichuan and Shandong	NGS, PCR
Sweet potato leaf curl Hubei virus*	Begomovirus	MH577011, MK951974, MK931305, MK931304	Hebei, Hubei, Hunan, Jiangsu, Shandong and Yunnan	NGS, PCR, RCA
Sweet potato leaf curl Canary virus	Begomovirus	MK931306	Guangdong, Hebei and Henan	PCR
Sweet potato leaf curl Georgia virus	Begomovirus	MK931310	Fujian, Hebei, Henan, Hainan, Shandong, Sichuan, Jiangxi and Zhejiang	PCR
Sweet potato leaf curl Guangxi virus	Begomovirus	MK931313, MK951966	Guangxi, Guangdong, Hebei, Henan, Hunan, Jiangsu, Shandong and Sichuan	PCR
Sweet potato leaf curl Sichuan virus 1	Begomovirus	MN186254	Guangdong, Guangxi, Henan, Hunan, Jiangsu and Sichuan	PCR
Sweet potato leaf curl Sichuan virus 2	Begomovirus	MK931315	Guangdong, Hebei, Henan, Hunan, Jiangsu and Sichuan	PCR
Sweet potato leaf curl virus	Begomovirus	MK951979, MK931320, MK931321	Anhui, Beijing, Chongqing, Fujian, Guangdong, Guangxi, Hainan, Hebei, Henan, Hubei, Hunan, Jiangsu, Jiangxi, Jilin, Shaanxi, Shandong, Sichuan and Yunnan	PCR
Sweet potato leaf curl China virus	Begomovirus	MK951977	Guangdong, Guizhou, Henan, Jiangsu, and Shandong	PCR
Cucumber mosaic virus	Cucumovirus	MK778778	Anhui, Chongqing, Fujian, Guangdong, Guangxi, Hainan, Henan, Hubei, Jiangsu, Shaanxi, Shandong, Shanxi, Sichuan and Zhejiang	NGS
Sweet potato collusive virus†	Cavemovirus	MK802082	Chongqing and Hainan	NGS, PCR
Tobacco mosaic virus†	Tobamovirus	MN186255	Henan	NGS
Sweet potato badnavirus A	Badnavirus	KJ463420	Anhui, Chongqing, Guangxi, Hebei, Henan, Hubei, Hunan, Jiangsu, Shaanxi and Shandong	NGS, PCR
Sweet potato badnavirus B	Badnavirus	KP407560	Henan, Hubei, Hunan, Jiangsu, Shaanxi and Shandong	NGS, PCR
Sweet potato symptomless virus 1	Mastrevirus	MG603668	Chongqing, Guangdong, Guangxi, Hebei, Henan, Jiangsu, Shandong and Shaanxi	NGS, PCR

Table 1 (continued)

Species	Genus	GenBank no. of representative sequences	Distribution (provinces)	Methods
Sweet potato chlorotic fleck virus	<i>Carlavirus</i>	MK778783	Anhui, Chongqing, Fujian, Guangdong, Guangxi, Guizhou, Hainan, Henan, Hunan, Jiangsu, Shaanxi, Shanxi, Shandong, Yunnan and Sichuan	NGS, PCR
Sweet potato chlorotic stunt virus	<i>Crinivirus</i>	MK802071	Anhui, Chongqing, Fujian, Guangdong, Guangxi, Guizhou, Hainan and Hebei, Henan, Hubei, Hunan, Jiangsu, Ningxia, Shaanxi, Shandong, Shanxi, Sichuan and Zhejiang	NGS, PCR

*New species; †First report of a sweet potato virus in China

nt sequence and deduced aa sequence identities among the B phytogroup isolates are 98.3–100% and 98.9–99.9%, respectively. The rest three isolates, 19-2, 32-1 and 33-1, were grouped together and were phylogenetically distant from the A and B phytogroups, suggesting that these three isolates belong to the C phytogroup.

The phylogenetic analysis according to polyprotein nt sequences revealed that the 18 SPVG isolates obtained were divided into five distinct phytogroups (Fig. 1 and Additional file 2: Figure S2). Pairwise comparisons of the polyprotein gene sequences of these 18 isolates revealed nt and deduced aa sequence identities in the range of 75.8–100% and 85.2–100%, respectively. Among them, nine isolates (CH-8-1, CH-31-2, CH-32-1, CH-ji-17-49, CH-L8, CH-L11, CH-S8, CH-xiang-17-2 and CH-yue-17-9) were grouped with the CH strain isolates (CH-shaan-15-3, MK778823 and CH-Jesus Maria, JQ824374); two isolates (CH2-32-1 and CH2-su-17-8) were grouped with the CH2 strain (CH2-shaan-15-3, MK778833 and CH2-WT325, KF790759); two isolates (CH3-8-1 and CH3-e-17-22) were clustered together with the CH3-66Al isolate (KX279878), with nt and aa sequence identities of 98.8–99.1% and 99.1–99.3%, respectively. CH3-8-1 and CH3-e-17-22 are phylogenetically distant from the CH and CH2 strain isolates, sharing nt and aa sequence identities of 82.7–83.6% and 91.2–91.9%, respectively, with the CH strain isolates, and 77.8–78.1% and 87.6–87.9%, respectively, with the CH2 strain isolates. Therefore, they were tentatively designated as SPVG-CH3 phytogroup isolates; one isolate (CH4-33-1) is phylogenetically distant from the CH and CH2 strain isolates, and also the CH3 phytogroup isolates, sharing nt and aa sequence identities of 79.5–80.6% and 89.5–90.2%, respectively, with the CH and CH2 strain isolates, and the CH3 phytogroup isolates; the remaining four isolates (CH5-8-1, CH5-gan-17-1, CH5-su-17-8 and CH5-yu-17-47) were clustered into a distant phytogroup, sharing nt and aa sequence identities in the range of

75.8–78.4% and 85.1–87.6%, respectively, with other isolates.

Pairwise comparisons of the polyprotein gene sequences of 10 SPLV isolates obtained revealed nt and deduced aa sequence identities in the range of 82.0–99.4% and 86.9–99.7%, respectively. The nt and aa sequence identities of these 10 isolates with five known isolates of SPLV (HG181, HN76, SC5, Taiwan and UN202) are 85.0–98.2% and 87.4–98.8%, respectively (Additional file 1: Table S3). A comparison of genomic sequences of the three SPV2 isolates (gan-17-1, lu-17-14 and su-17-8) that we obtained and 10 known SPV2 isolates, namely, GJ118, AM-MB2, AuScan, CW142, GWB-2, HN77, LSU2, SC6, SCN20 and TM37, presented nt and aa sequence identities of 97.8–98.7% and 97.9–98.9%, respectively (Additional file 1: Table S3). This suggests that the genome sequences of SPV2 are highly conserved. In addition, the polyprotein nt and aa sequence identities among the 11 SPVC isolates obtained are 87.8–100% and 90.1–100%, respectively. These 11 SPVC isolates presented 88.2–99.0% and 90.4–99.3% sequence identities, respectively, with three known isolates (Bungo, C1 and Ch1) (Additional file 1: Table S3).

Genetic diversity of sweepoviruses

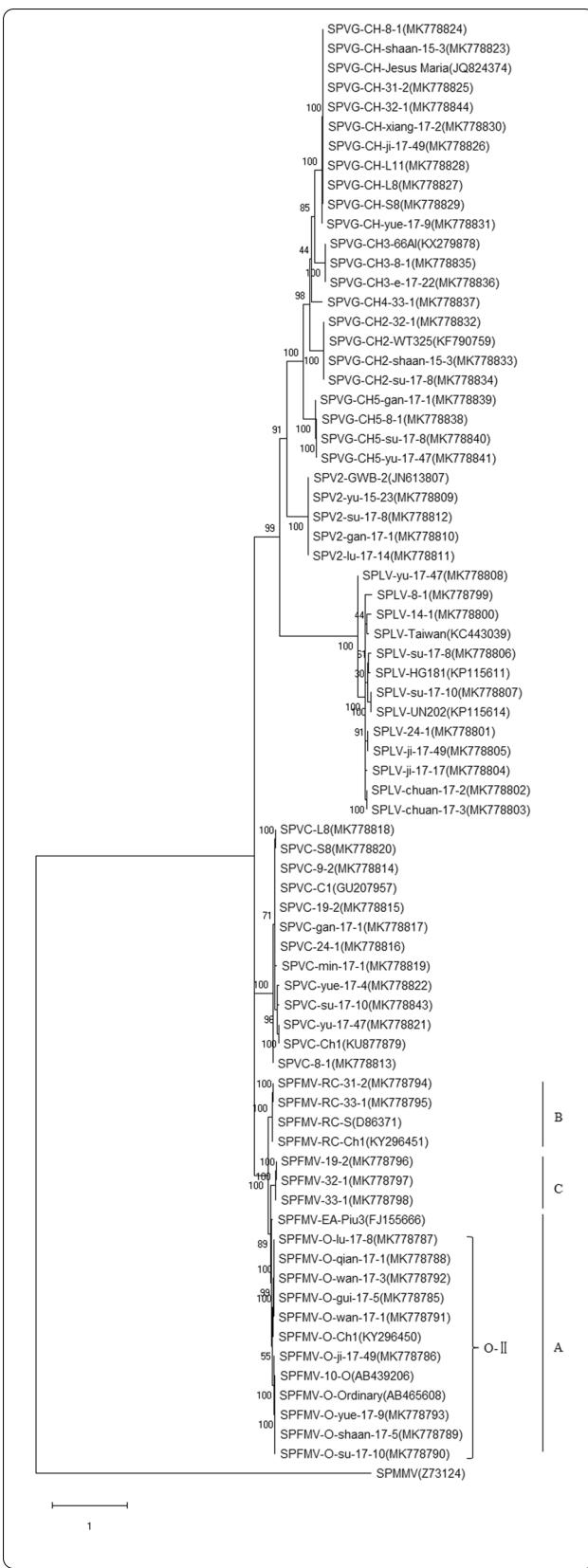
Full-length genomic sequences of 40 sweepovirus isolates from 12 provinces (Additional file 1: Table S2) were obtained, with the nt sequence identity among these isolates ranging from 68.0 to 99.8%. According to the criteria set by the ICTV for begomovirus species demarcation (91% nt sequence identity) and the SDT v1.2 analysis result (Brown et al. 2015), 29 of these 40 isolates were classified into seven species: SPLCCV, SPLCGoV, SPL-CGV, SPLCSiV1, SPLCSiV2, SPLCV and SPLCCNV (Additional file 1: Tables S2, S4). A comparison of the complete genomic sequences of the remaining 11 isolates (MH577011, MK951969, MK951970, MK951971, MK951972, MK951973, MK951974, MK951975, MK931304, MK931305 and MK951976) exhibited the

Fig. 1 Phylogenetic tree of polyprotein nucleotide sequences of SPFMV, SPLV, SPVG, SPVC and SPV2 isolates. The tree was generated using MEGA X (the Maximum Likelihood method, General Time Reversible model and 1000 bootstrap replicates). Sweet potato mild mottle virus (SPMMV; Z73124) was used as an outgroup taxon

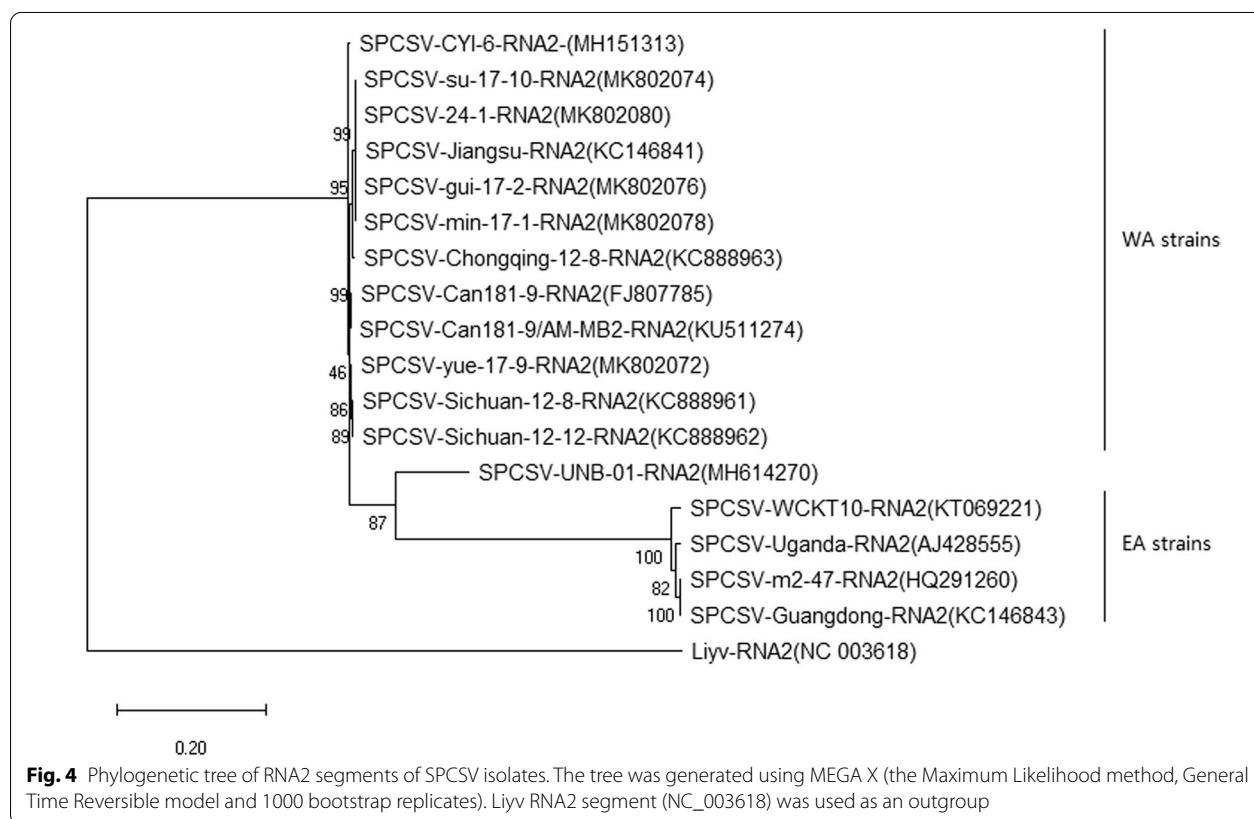
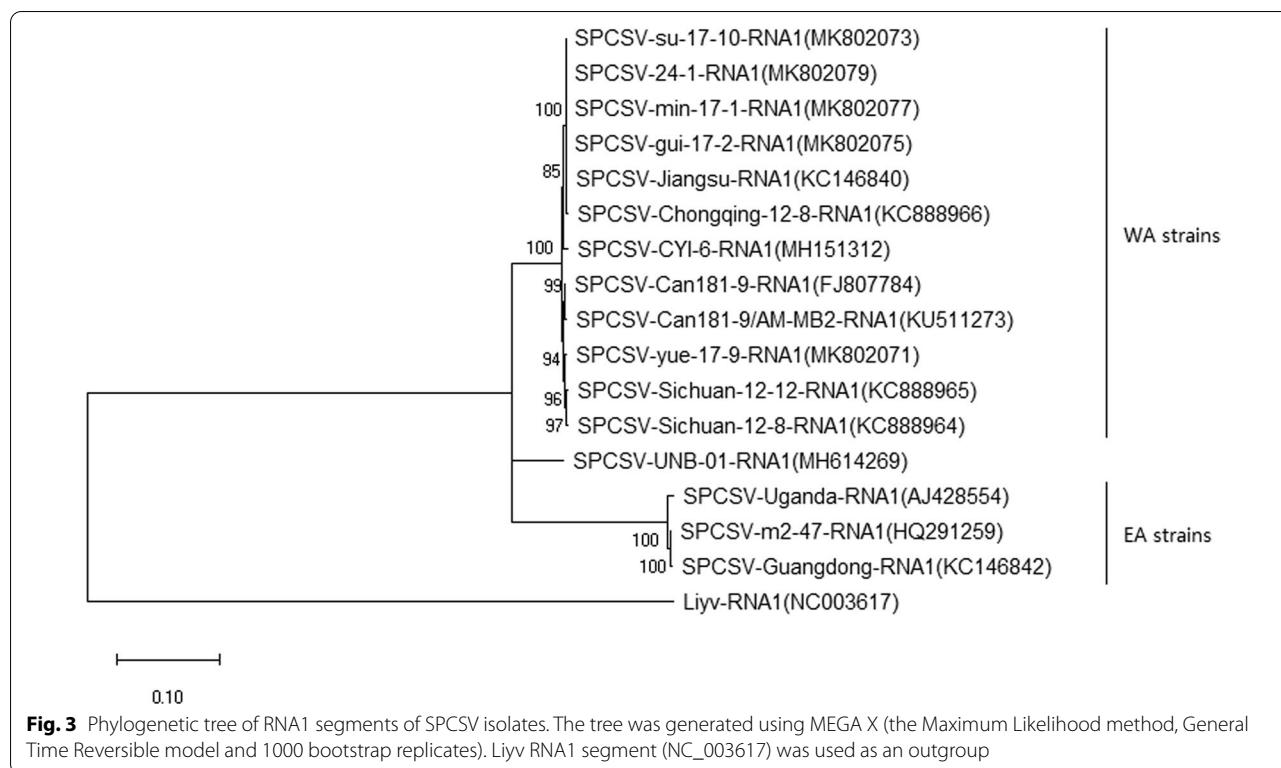
highest nt sequence identity, 73.4%, with known begomoviruses, indicating that the nt sequence identity was lower than 91% (Additional file 1: Table S4). Furthermore, the full-length genomic sequence identity among the 11 isolates ranged from 89.6% to 99.8%. Therefore, these 11 isolates belong to a same and novel species, with *Sweet potato leaf curl Hubei virus* (SPLCHbV) as the proposed name. According to the strain demarcation criteria for begomoviruses (ranges 91–94% nt sequence identity), as proposed by the ICTV (Haible et al. 2006), these 11 isolates of SPLCHbV are composed of four strains with SPLCHbV-Hb, SPLCHbV-Js, SPLCHbV-Hn and SPLCHbV-Sd as the proposed names (Fig. 2, Additional file 1: Table S5 and Additional file 2: Figure S3).

Complete genome sequence comparisons using SDT v1.2 revealed that the genomic sequence identity of the 6 SPLCGV isolates from 3 provinces ranged from 90.4 to 99.4%. Further phylogenetic analysis revealed that four (MK951965, MK951966, MK951967 and MK951968) of these isolates presented an nt sequence identity of 98.2–99.4% (Additional file 1: Table S6 and Additional file 2: Figure S4) and are phylogenetically distant from other SPLCGV isolates analyzed (Fig. 2). They share the highest identity of 93.1% with other known SPLCGV isolates, which is lower than the criteria required for begomoviruses strain classification (94% limit); therefore, these four isolates constitute a novel strain, designated as SPLCGV-Gd. In addition, the complete genomic sequences of 7 SPLCV isolates were obtained from 6 provinces, with their nt sequence identity ranging from 82.4 to 98.1%. The SDT and phylogenetic analyses revealed that these 7 isolates belong to strains SPLCV-US, SPLCV-KR and SPLCV-Fu (Fig. 2 and Additional file 1: Table S7). The genome sequences of 5 other sweepoviruses, namely, SPLCCV, SPLCGoV, SPLCSiV1, SPLCSiV2 and SPLCCNV, are conservative, with 94.8–99.7% nt sequence identity.

The 40 sweepovirus isolates showed typical genomic structural features of sweepoviruses: genome size ranging from 2770 to 2821 bp; six open read frames (ORFs); viral-sense strand with two ORFs (AV1 and AV2), and complementary-sense strand with four ORFs (AC1, AC2, AC3 and AC4). The genomes of these 40 sweepovirus isolates all encompass a highly conserved stem-loop structure containing the TAATATTAC/T sequence.







Molecular variation of SPCSV

Nearly full-length genomic sequences of 5 SPCSV isolates were obtained from 5 provinces (Additional file 1: Table S2). The RNA1 and RNA2 segments of the 5 isolates have 4 and 9 ORFs, respectively, similar to those of the Jiangsu isolate of the SPCSV WA strain (Qin et al. 2013b). The nt sequence identities of RNA1 and RNA2 among the five isolates are 98.9–100% and 98.8–99.9%, respectively. The phylogenetic analysis revealed that the five isolates belong to the WA strain (Figs. 3, 4). Therefore, the WA strain could be the dominant strain in sweet potatoes in China, and this is consistent with the findings of our previous study (Qin et al. 2013b).

Molecular variations in other viruses

Full-length or nearly full-length genome sequences of 2 cucumber mosaic virus (CMV) isolates, 2 SPCFV isolates, 1 TMV isolate and 5 SPSMV-1 isolates were obtained (Additional file 1: Table S2).

For CMV, segments RNA1, RNA2 and RNA3 in the lu-17-14 isolate presented the highest nt sequence identities of 98.0%, 98.7% and 98.4% with RNA1 (KT350980) and RNA2 (KT350981) of a maize isolate from China, and RNA3 (JX993913) of a Chinese thorowax root isolate from China, respectively. RNA1, RNA2 and RNA3 of the yang-SS3-2 isolate presented the highest nt sequence identity with RNA1 (AB179764) of a *Raphanus sativus* isolate from Japan, RNA2 (KX883764) of a true fly isolate from China, and RNA3 (EF213025) of a *Brassica chinensis* isolate from China, at 96.8%, 97.5% and 97.1%, respectively. Phylogenetic analysis based on the nt sequences of coat protein (CP) genes with known subgroup CMV isolates (Roossinck 2002; Bald-Blumea et al. 2017) showed that lu-17-14 and yang-SS3-2 belong to subgroup IB (Additional file 2: Figure S5).

Sequence analysis revealed that the genome of the two SPCFV isolates (gui-17-2-1 and wan-17-1) has six complete ORFs encoding RNA-dependent RNA polymerase (RdRp), triple gene block protein 1 (TGB1), TGB2, TGB3, CP and nucleic acid binding protein (NaBP), and exhibited the typical genomic structural features of carlaviruses (Deng et al. 2015). The nt sequence identity between these two isolates is 73.8%. The phylogenetic analysis results indicate that gui-17-2-1 and wan-17-1 are distant (Additional file 2: Figure S6), suggesting a high molecular variability within SPCFV genomic sequences.

Nearly full-length genomic sequence of one TMV isolate was obtained (Additional file 1: Table S2). The isolate has the highest nt sequence identity of 99.8% with a *Nicotiana benthamiana* isolate from Spain (MK087763). To the best of our knowledge, this is the first study to report TMV infecting sweet potatoes in China.

The full-length genomic sequences of 5 SPSMV-1 isolates were obtained (Additional file 1: Table S2). Sequence comparisons indicate that the nt sequence identity of these 5 isolates with other isolates of the virus in GenBank ranged from 97.4 to 99.8% (Additional file 1: Table S8 and Additional file 2: Figures S7), suggesting that the SPSMV-1 genomic sequences are highly conserved.

Discussion

In the present study, viruses infecting sweet potato in China were identified using NGS, PCR and RCA approaches. Twenty-one virus species were identified, including 1 novel virus and 5 novel viral strains, and 2 viruses were found to infect sweet potato in China for the first time. The results highlight the high diversity of the sweet potato viromes in China. The full-length or nearly full-length genomic sequences of 111 isolates were obtained, belonging to 18 sweet potato virus species, and the molecular diversity of major sweet potato viruses was analyzed. The present study for the first time comprehensively reports the variability of sweet potato viruses in China.

Genomic sequence comparisons indicate that three SPF MV isolates (19-2, 32-1 and 33-1) obtained in this study share nt and aa sequence identities of 86.0–90.7% and 92.3–95.8% with the O, RC and EA strain isolates, respectively. Previous studies have reported that the polyprotein nt and aa sequence identities between different SPF MV strains are lower than 94.0% and 96.0%, respectively (Adams et al. 2005; Yamasaki et al. 2010; Wylie et al. 2017). Therefore, these three isolates could be a novel SPF MV strain, and further research on biological characteristics of these three isolates is warranted.

The detection of eight sweepoviruses in sweet potatoes in China is the highest number of virus species detected in a study to date in the country. The eight sweepoviruses include one novel species, which was identified in the present study, and four novel species recently identified in China (Luan et al. 2006; Liu et al. 2013, 2014, 2017). The findings indicate that there is a great diversity in sweepoviruses found on sweet potatoes in China. Sweepoviruses are transmitted by the whitefly (*Bemisia tabaci*), an important agricultural pest transmitting numerous economically important plant viruses, which has recently exhibited increasing trends in sweet potato cultivation regions in China (Gilbertson et al. 2015). The diversity of sweepovirus could be associated with the activities of whiteflies on sweet potatoes in China. SPCSV is another whitefly-transmitted virus. In the present study, SPCSV was detected from 18 provinces. SPCSV was reported in Guangdong Province of China for the first time in 2011 (Qiao et al. 2011). The rapid spread of SPCSV could also be associated with the expansion of

whitefly populations in China. Therefore, controlling whitefly populations in sweet potato fields is a potential strategy to manage SPCSV incidence and proliferation. SPCSV can also synergistically infect sweet potatoes with several other viruses (Karyeija et al. 2000; Mukasa et al. 2006; Cuellar et al. 2011, 2015). Co-infection by SPCSV and SPFMV causes SPVD, a destructive disease in sweet potatoes (Schaefers and Terry 1976; Clark et al. 2012). According to the NGS results, SPVD detection rate was as high as 48.4%, suggesting that SPVD is a common disease (Additional file 1: Table S9) and poses a major threat to sweet potato production in China. Therefore, it is vital to bolster SPVD monitoring and control activities.

In the present study, sweet potato samples were collected randomly from sweet potato fields in 25 provinces, where the areas under sweet potato cultivation vary considerably. The amount of sweet potato plants sampled was not proportional to sweet potato cultivation area in the provinces. Although as many as 21 virus species were identified in the present study, their distribution and incidence remain unclear in China. Consequently, more samples should be collected and more reliable sampling efforts should be implemented in future studies. Nonetheless, our results provide an important basis for research on the control and management of sweet potato viral diseases in China.

Conclusions

We identified 21 virus species, including 12 DNA and 9 RNA viruses, in the sweet potato samples from China. One novel sweepovirus, SPLCHbV, was identified. Two species, SPCV and TMV, were identified for the first time in sweet potato in China. Genome sequence comparisons of five potyvirus indicate that the genome of SPV2 is highly conserved, and the other four potyviruses, SPFMV, SPVG, SPLV and SPVC, exhibited a high genetic variability. The similarity among the sweepovirus genomic sequences obtained from eight sweepovirus species varied greatly. The genome sequences of SPCSV are highly conserved. The present study for the first time comprehensively reports the variability of sweet potato viruses in China. The results provide a basis for future research on the control and management of sweet potato viral diseases in China.

Methods

Collection of samples

Approximately 2000 samples with viral disease-like symptoms such as leaf shriveling, curling, malformation, vein clearing, chlorosis and mosaic were randomly collected from sweet potato fields in 25 provinces in China during 2014–2019. Most of the samples were leaves, and

only a few were storage root and seed samples. The collected leaf and storage root samples were stored at –80 °C until further analyses, and the seed samples were stored under dry conditions at room temperature until use.

Primers

Primers for virus detection and viral genome sequence amplification were designed based on the nucleotide sequences of sweet potato viruses obtained from GenBank or NGS-assembled sequences (Additional file 1: Tables S10, S11). All primers were synthesized by Sangon Bioengineering and Technology Service Co., Ltd. (Shanghai, China).

Nucleic acid extraction

DNA and RNA were extracted from samples ground in liquid nitrogen, using the DNA extraction Kit (Omega Bio-tek, Norcross, USA) and the Total Plant RNA Extraction Miniprep System (Sangon, Shanghai, China) according to the manufacturer's instructions. The extracted nucleic acid samples were stored at –80 °C until use.

Next-generation sequencing and data processing

Sixty-four samples were used for NGS (Additional file 1: Table S9). The total RNA was extracted from sweet potato samples using the TRIzol Kit (Invitrogen, Carlsbad, CA, USA), and its concentration and purity was measured using the Agilent 2100 Bioanalyzer (Agilent Technologies, Palo Alto, CA) and by agarose gel electrophoresis. Subsequently, the small RNAs were enriched using the sRNAeasy kit (Qiagen, Dusseldorf, Germany) to construct small RNA libraries for small RNA sequencing (sRNA-seq); the sRNA libraries were built using the TruSeq Small RNA Sample Prep Kit (Illumina, San Diego, USA). An Illumina Hiseq2500 platform (Illumina) set with 50-bp read lengths was used for sequencing (BGI Tech Company, Shenzhen, China). The ribosome total RNA was removed using the RiboZero Magnetic Kit (Epicenter, USA) to construct ribo-depleted RNA libraries for RNA sequencing (RNA-Seq). The libraries were built using the TruSeq RNA Sample Prep Kit (Illumina). An Illumina HiSeq X-ten platform (Illumina) set with 150-bp pair-end reads was used for sequencing (BGI Tech Company). After deep sequencing, adaptor and low-quality sequences were removed from raw reads, using Trimmomatic (Bolger et al. 2014), yielding more than 8-Gb clean reads for each sample library. The remaining RNA-Seq reads were mapped to the genome sequences of sweet potato (downloaded from <http://public-genomes/ngs.molgen.mpg.de/SweetPotato/>) (Yang et al. 2017) using CLC Genomic Workbench 10.0 (Qiagen). The reads with sequence similarities of > 60% to the sweet potato genome

sequences were eliminated to reduce interference of the host background, and the remaining unique reads were de novo assembled using CLC Genomic Workbench 10.0 (Qiagen). The clean reads of 18–26 nt from sRNA-Seq were subjected to de novo assembly using CLC Genomic Workbench 10.0 (Qiagen). The resulting contigs were subjected to BLASTx and BLASTn searches against viral (taxid:10239) and viroidal (taxid:2559587) sequences of local datasets retrieved from the National Center for Biotechnology Information (NCBI) databank (Wu et al. 2015; Zhang et al. 2021). These processes enabled the identification of contigs with viral sequence attributes.

Rolling-circle amplification

One hundred samples with leaf curl symptoms were used in rolling-circle amplification (RCA). DNA extracted from the samples was amplified using the TempliPhi™ 100 Amplification Kit (GE Healthcare, USA) according to the method of Haible et al. (2006). The RCA products (5 µL) were digested in a 30-µL container using the restriction enzyme *Bam*HI (10 U) (TaKaRa, Japan) at 37 °C for 4 h. The digested products were separated on 1% agarose gel. The DNA fragments (2.8 kb) were purified using the Gel Extraction Kit (Axygen, USA), ligated into the PUC-118 vector, and then transformed into *Escherichia coli* strain JM109. Positive clones were identified by PCR and sequenced by TaKaRa Bioengineering Co., Ltd. (Dalian, China).

(RT-)PCR and sequencing

For RNA viruses, the extracted RNA samples were reverse transcribed into cDNA using M-MLV reverse transcriptase (TaKaRa) and reverse primers of the corresponding viral species (Additional file 1: Table S10), followed by PCR amplification with the synthesized cDNA as the template. The PCR mixture (25 µL) consisted of 2 µL of cDNA, 2.5 µL of 10×Ex *Taq* or LA *Taq* Buffer ($MgCl_2$), 2 µL of 2.5 mM dNTP mixture, 2 µL of each forward and reverse primers (Additional file 1: Table S10), and 0.2 µL of Ex *Taq* DNA polymerase or LA *Taq* DNA polymerase. PCR amplifications were performed under the following conditions: an initial denaturation for 5 min at 94 °C, followed by 35 cycles at 94 °C for 1 min, 55 °C for 1 min, 72 °C for 1 min/kb, and a final extension at 72 °C for 10 min. The PCR products were separated on 1% agarose gels for 30 min. Fragments of interest were recovered using the Gel Extraction Kit (Omega Bio-tek) according to the manufacturer's recommendation. The eluted DNA was ligated into the pMD19-T vector (TaKaRa) according to the

manufacturer's instructions and cloned into *E. coli* strain JM109. Plasmids containing the amplified viral sequence were sequenced by TaKaRa Bioengineering Co., Ltd.. For DNA viruses, DNA was extracted and used as the template for PCR amplification. The PCR mixture, cycling conditions and sequencing procedures were similar to those for RNA viruses.

Cloning and sequencing of the genome of sweet potato viruses

Full-length genomic sequences of potyviruses were cloned by subsection amplification, and the primers used are listed in Additional file 1: Table S11. RT-PCR was performed as previously described. Overlapping fragments of 500–1000 bp were amplified and sequenced after cloning. At least three independent clones for each fragment were sequenced. 5'-RACE and 3'-RACE were conducted by TaKaRa Bioengineering Co., Ltd. Full-length genomic sequences of sweepoviruses and SPSMV-1 were cloned using the back-to-back primers (Additional file 1: Table S11) or RCA. The genomic sequences of other viruses, including TMV, CMV, SPCFV and SPCSV were obtained by NGS assembly.

Sequence analysis

Sequence analysis and comparisons were performed using DNAMAN v8.1 (Lynnon Biosoft, Quebec), Sequence Demarcation Tool (SDT) v1.2 (Muhire et al. 2014) and BLAST (NCBI). A phylogenetic tree was constructed using the maximum likelihood (ML) method with MEGA X v10.1 (Kumar et al. 2018). The obtained viral nucleotide sequences and the reference sequences are listed in Additional file 1: Tables S1, S2, respectively.

Abbreviations

CMV: Cucumber mosaic virus; CP: Coat protein; GLV: Garlic latent virus; ICTV: International Committee on Taxonomy of Viruses; MSV: Maize streak virus; NaBP: Nucleic acid binding protein; NCBI: National Center for Biotechnology Information; NGS: Next-generation sequencing; ORFs: Open read frames; PSV: Peanut stunt virus; RCA: Rolling-circle amplification; RdRp: RNA-dependent RNA polymerase; SDT: Sequence demarcation tool; SPBV-A: Sweet potato badnavirus A; SPBV-B: Sweet potato badnavirus B; SPCV: Sweet potato colloidal virus; SPCFV: Sweet potato chlorotic fleck virus; SPCSV: Sweet potato chlorotic stunt virus; SPFMV: Sweet potato feathery mottle virus; SPLCCNV: Sweet potato leaf curl China virus; SPLCCV: Sweet potato leaf curl Canary virus; SPLCGoV: Sweet potato leaf curl Georgia virus; SPLCGV: Sweet potato leaf curl Guangxi virus; SPLCHbV: Sweet potato leaf curl Hubei virus; SPLChNv: Sweet potato leaf curl Henan virus; SPLCsDv: Sweet potato leaf curl Shandong virus; SPLCSIV1: Sweet potato leaf curl Sichuan virus 1; SPLCSIV2: Sweet potato leaf curl Sichuan virus 2; SPLCV: Sweet potato leaf curl virus; SPLV: Sweet potato latent virus; SPMMV: Sweet potato mild mottle virus; SPSMV-1: Sweet potato symptomless virus 1; SPV2: Sweet potato virus 2; SPVC: Sweet potato virus C; SPVD: Sweet potato virus disease; SPVG: Sweet potato virus G; TGB1: Triple gene block protein 1; TMV: Tobacco mosaic virus.

Supplementary Information

The online version contains supplementary material available at <https://doi.org/10.1186/s42483-021-00097-8>.

Additional file 1: Table S1. Virus species detected in sweet potato fields in different provinces. **Table S2.** Virus isolates obtained in the present study. **Table S3.** Reference sequences used in the present study. **Table S4.** SDT analysis of sweepoviruses. **Table S5.** SDT analysis of SPLCHbV. **Table S6.** SDT analysis of SPLCGV. **Table S7.** SDT analysis of SPLCV. **Table S8.** SDT analysis of SPSMV-1. **Table S9** Viruses detected from 64 samples by next-generation sequencing. **Table S10.** Primers used for virus detection in the present study. **Table S11.** Primers used for complete genome amplification in the present study.

Additional file 2: Figure S1. Phylogenetic tree reconstructed using the polyprotein nucleotide sequences of the SPFMV isolates. The tree was generated using MEGA X (MUSCLE alignment, neighbor-joining and p-distance model, and 1000 bootstrap replicates). Sweet potato mild mottle virus (SPMMV) (Z73124) was used as an outgroup taxon. **Figure S2.** Phylogenetic trees reconstructed using the polyprotein nucleotide sequences of the SPVG isolates. The tree was generated using MEGA X (MUSCLE alignment, neighbor-joining and p-distance model, and 1000 bootstrap replicates). Sweet potato mild mottle virus (SPMMV) (Z73124) was used as an outgroup taxon. **Figure S3.** Phylogenetic tree illustrating the relationship among 9 SPLCHbV sequences. The tree was generated using MEGA X (MUSCLE alignment, neighbor-joining and p-distance model, and 1000 bootstrap replicates). **Figure S4.** Phylogenetic tree illustrating the relationship among 8 SPLCGV sequences. The tree was generated using MEGA X (MUSCLE alignment, neighbor-joining and p-distance model, and 1000 bootstrap replicates). **Figure S5.** Phylogenetic tree based on the nucleotide sequences of CP of CMV isolates. The tree was generated using MEGA X (MUSCLE alignment, neighbor-joining and p-distance model, and 1000 bootstrap replicates). Peanut stunt virus (PSV) (U15730) was used as an outgroup taxon. **Figure S6.** Phylogenetic tree constructed using near complete genome sequences of SPCFV isolates. The tree was generated using MEGA X (MUSCLE alignment, neighbor-joining and p-distance model, and 1000 bootstrap replicates). Garlic latent virus (GLV) (NC_003557) was used as an outgroup taxon. **Figure S7.** Phylogenetic tree constructed using the complete genome sequences of SPSMV-1 isolates. The tree was generated using MEGA X (MUSCLE alignment, neighbor-joining and p-distance model, and 1000 bootstrap replicates). Maize streak virus (MSV) (HQ693373) was used as an outgroup taxon.

Acknowledgements

We thank Dr. Cao Mengji from Southwest University, China for conducting partial NGS analysis and critically reading the manuscript.

Authors' contributions

ZCZ conceived the project. YJW, YHQ and QQ performed most of the experiments with assistance from SW, DSZ, YTT, FMZ, YZW and HL. All authors analyzed the data. ZCZ, QQ, YJW and YHQ wrote the manuscript. All authors read and approved the final manuscript.

Funding

This work was supported by China Agriculture Research System of MOF and MARA (CARS-10-B13), the project from Industry-University-Research Cooperation of Henan Province of China (172107000038) and the Science-Technology Foundation for Outstanding Young Scientists of Henan Academy of Agricultural Sciences (Grant No. 2018YQ19).

Availability of data and materials

Not applicable.

Declarations

Ethics approval and consent to participate

Not applicable.

Consent for publication

Not applicable.

Competing interests

The authors declare that they have no competing interests.

Author details

¹Institute of Plant Protection, Henan Academy of Agricultural Sciences, Zhengzhou, Henan, China. ²IPM Key Laboratory in Southern Part of North China for Ministry of Agriculture, Zhengzhou, Henan, China.

Received: 18 May 2021 Accepted: 11 August 2021

Published online: 30 August 2021

References

- Adams MJ, Antoniw JF, Fauquet CM. Molecular criteria for genus and species discrimination within the family Potyviridae. *Arch Virol*. 2005;150:459–79. <https://doi.org/10.1007/s00705-004-0440-6>.
- Aritua V, Barg E, Adipala E, Gibson RW, Lesemann DE, Vetten HJ. Host range, purification, and genetic variability in sweet potato chlorotic fleck virus. *Plant Dis*. 2009;93(1):87–93. <https://doi.org/10.1094/PDIS-93-1-0087>.
- Ateka EM, Barg E, Njeru RW, Thompson G, Vetten HJ. Biological and molecular variability among geographically diverse isolates of sweet potato virus 2. *Arch Virol*. 2007;152:479–88. <https://doi.org/10.1007/s00705-006-0879-8>.
- Bald-Blumea N, Bergervoet JHW, Maissa E. Development of a molecular assay for the detection of cucumber mosaic virus and the discrimination of its subgroups I and II. *J Virol Methods*. 2017;243:35–43. <https://doi.org/10.1016/j.jviromet.2017.01.011>.
- Bolger AM, Lohse M, Usadel B. Trimmomatic: a flexible trimmer for Illumina sequence data. *Bioinformatics*. 2014;30(15):2114–20. <https://doi.org/10.1093/bioinformatics/btu170>.
- Brown JK, Zerbini FM, Navas-Castillo J, Moriones E, Ramos-Sobrinho R, Silva JCF, et al. Revision of *Begomovirus* taxonomy based on pairwise sequence comparisons. *Arch Virol*. 2015;160:1593–619. <https://doi.org/10.1007/s00705-015-2398-y>.
- Clark CA, Davis JA, Abad JA, Cuellar WJ, Fuentes S, Kreuze JF, et al. Sweet potato viruses: 15 years of progress on understanding and managing complex diseases. *Plant Dis*. 2012;96(2):168–85. <https://doi.org/10.1094/PDIS-07-11-0550>.
- Colinet D, Kummert J. Identification of a sweet potato feathery mottle virus isolate from China (SPFMV-CH) by the polymerase chain reaction with degenerate primers. *J Virol Methods*. 1993;45(2):149–59. [https://doi.org/10.1016/0166-0934\(93\)90099-D](https://doi.org/10.1016/0166-0934(93)90099-D).
- Colinet D, Kummert J, Lepoivre P. The complete nucleotide sequences of the coat protein cistron and the 3' non-coding region of a newly-identified potyvirus infecting sweetpotato, as compared to those of sweetpotato feathery mottle virus. *Arch Virol*. 1994;139:327–36. <https://doi.org/10.1007/BF01310795>.
- Colinet D, Kummert J, Lepoivre P. Evidence for the assignment of two strains of SPLV to the genus *Potyvirus* based on coat protein and 3' non-coding region sequence data. *Virus Res*. 1997;49(1):91–100. [https://doi.org/10.1016/S0168-1702\(97\)01456-1](https://doi.org/10.1016/S0168-1702(97)01456-1).
- Colinet D, Nguyen M, Kummert J, Lepoivre P, Xia FZ. Differentiation among potyviruses infecting sweet potato based on genus- and virus-specific reverse transcription polymerase chain reaction. *Plant Dis*. 1998;82(2):223–9. <https://doi.org/10.1094/PDIS.1998.82.2.223>.
- Cuellar WJ, De Souza J, Barrantes I, Fuentes S, Kreuze JF. Distinct cavemoviruses interact synergistically with sweet potato chlorotic stunt virus (genus *Crinivirus*) in cultivated sweet potato. *J Gen Virol*. 2011;92(5):1233–43. <https://doi.org/10.1099/vir.0.029975-0>.
- Cuellar WJ, Galvez M, Fuentes S, Tugume J, Kreuze J. Synergistic interactions of begomoviruses with sweet potato chlorotic stunt virus (genus *Crinivirus*) in sweet potato (*Pomoea batatas* L.). *Mol Plant Pathol*. 2015;16(5):459–71. <https://doi.org/10.1111/mpp.12200>.
- de Albuquerque TMR, Sampaio KB, de Souza EL. Sweet potato roots: unrevealing an old food as a source of health promoting bioactive compounds—a review. *Trends Food Sci Technol*. 2019;85:277–86. <https://doi.org/10.1016/j.tifs.2018.11.006>.

- Deng XG, Peng XJ, Zhu F, Chen YJ, Zhu T, Qin SB, et al. A critical domain of sweet potato chlorotic fleck virus nucleotide binding protein (NaBp) for RNA silencing suppression, nuclear localization and viral pathogenesis. *Mol Plant Pathol.* 2015;16(4):365–75. <https://doi.org/10.1111/mpp.12186>.
- FAOSTAT. 2019. Available from: <http://faostat.fao.org/>. Accessed date: 07/23/2021.
- Gilbertson RL, Batuman O, Webster CG, Adkins S. Role of the insect supervectors *Bemisia tabaci* and *Frankliniella occidentalis* in the emergence and global spread of plant viruses. *Annu Rev Virol.* 2015;2:67–93. <https://doi.org/10.1146/annurev-virology-031413-085410>.
- Gu YH, Tao X, Lai XJ, Wang HY, Zhang YZ. Exploring the polyadenylated RNA virome of sweet potato through high-throughput sequencing. *PLoS ONE.* 2014;9(6):e98884. <https://doi.org/10.1371/journal.pone.0098884>.
- Hadidi A, Flores R, Candresse T, Barba M. Next-generation sequencing and genome editing in plant virology. *Front Microbiol.* 2016;7:1325. <https://doi.org/10.3389/fmicb.2016.01325>.
- Haible D, Kober S, Jeske H. Rolling circle amplification diagnosis and genomics of geminiviruses. *J Virol Methods.* 2006;135(1):9–16. <https://doi.org/10.1016/j.jviromet.2006.01.017>.
- Hassan I, Orílio AF, Fiallo-Olivé E, Briddon RW, Navas-Castillo J. Infectivity, effects on helper viruses and whitefly transmission of the deltasatellites associated with sweepoviruses (genus *Begomovirus*, family *Geminiviridae*). *Sci Rep.* 2016;6:30204. <https://doi.org/10.1038/srep30204>.
- Karveija RF, Kreuze JF, Gibson RW, Valkonen JPT. Synergistic interactions of a potyvirus and a phloem-limited crinivirus in sweet potato plants. *Virology.* 2000;269(1):26–36. <https://doi.org/10.1006/viro.1999.0169>.
- Kreuze JF, Perez A, Untiveros M, Quispe D, Fuentes S, Barker I, et al. Complete viral genome sequence and discovery of novel viruses by deep sequencing of small RNAs: a generic method for diagnosis, discovery and sequencing of viruses. *Virology.* 2009;388(1):1–7. <https://doi.org/10.1016/j.virol.2009.03.024>.
- Kumar S, Stecher G, Li M, Knyaz C, Tamura K. MEGA X: Molecular evolutionary genetics analysis across computing platforms. *Mol Biol Evol.* 2018;35(6):1547–9. <https://doi.org/10.1093/molbev/msy096>.
- Li XC, Zhang ZC, Qiao Q, Qin YH, Wang S, Zheng WM. Molecular detection and expression of the coat protein gene of sweet potato badnavirus B in Escherichia coli. *J Plant Prot.* 2016;43(2):345–6 (in Chinese) <https://doi.org/10.13802/j.cnki.zwbhxb.2016.02.025>.
- Liu Q, Zhang Z, Qiao Q, Qin Y, Zhang D, Tian Y, et al. Complete genome sequence of a novel monopartite begomovirus infecting sweet potato in China. *Virus Genes.* 2013;47:591–4. <https://doi.org/10.1007/s11262-013-0982-3>.
- Liu Q, Zhang Z, Li J, Qiao Q, Qin Y, Zhang D, et al. Complete genome sequence of a novel monopartite begomovirus infecting sweet potato in China. *Arch Virol.* 2014;159:1537–40. <https://doi.org/10.1007/s00705-013-1958-2>.
- Liu Q, Wang Y, Zhang Z, Lv H, Qiao Q, Qin Y, et al. Diversity of sweepoviruses infecting sweet potato in China. *Plant Dis.* 2017;101(12):2098–103. <https://doi.org/10.1094/PDIS-04-17-0524-RE>.
- Luan YS, Zhang J, An LJ. First report of sweet potato leaf curl virus in China. *Plant Dis.* 2006;90(8):1111. <https://doi.org/10.1094/PD-90-1111C>.
- Luan YS, Zhang J, Liu DM, Li WL. Molecular characterization of sweet potato leaf curl virus isolate from China (SPLCV-CN) and its phylogenetic relationship with other members of the Geminiviridae. *Virus Genes.* 2007;35:379–85. <https://doi.org/10.1007/s11262-007-0084-1>.
- Muhire BM, Varsani A, Martin DP. SDT: a virus classification tool based on pairwise sequence alignment and identity calculation. *PLoS ONE.* 2014;9(9):e108277. <https://doi.org/10.1371/journal.pone.0108277>.
- Mukasa SB, Rubaihayo PR, Valkonen JPT. Interactions between a crinivirus, an ipomovirus and a potyvirus in coinfecting sweetpotato plants. *Plant Pathol.* 2006;55(3):458–67. <https://doi.org/10.1111/j.1365-3059.2006.01350.x>.
- Qiao Q, Zhang ZC, Qin YH, Zhang DS, Tian YT, Wang YJ. First report of sweet potato chlorotic stunt virus infecting sweet potato in China. *Plant Dis.* 2011;95(3):356. <https://doi.org/10.1094/PDIS-09-10-0675>.
- Qin Y, Zhang Z, Qiao Z, Qiao Q, Zhang D, Tian Y, et al. First report of sweet potato leaf curl Georgia virus on sweet potato in China. *Plant Dis.* 2013a;97(10):1388. <https://doi.org/10.1094/PDIS-10-12-0967-PDN>.
- Qin Y, Zhang Z, Qiao Q, Zhang D, Tian Y, Wang Y, et al. Complete genome sequences of two sweet potato chlorotic stunt virus isolates from China. *Genome Announc.* 2013b;1(3):e00218-e313. <https://doi.org/10.1128/genomeA.00218-13>.
- Qin YH, Li XC, Zhang ZC, Qiao Q, Zhang DS, Wang YJ, et al. First report of sweet potato badnavirus A in China. *Plant Dis.* 2016;100(4):865. <https://doi.org/10.1094/PDIS-09-15-1081-PDN>.
- Roossinck MJ. Evolutionary history of cucumber mosaic virus deduced by phylogenetic analyses. *J Virol.* 2002;76(7):3382–7. <https://doi.org/10.1128/JVI.76.7.3382-3387.2002>.
- Schaefers GA, Terry ER. Insect transmission of sweetpotato disease agents in Nigeria. *Phytopathology.* 1976;66:642–5. <https://doi.org/10.1094/Phyto-66-642>.
- Wang YJ, Zhang DS, Zhang ZC, Wang S, Qiao Q, Qin YH, et al. First report on sweetpotato symptomless virus 1 (genus *Mastrevirus*, family *Geminiviridae*) in sweetpotato in China. *Plant Dis.* 2015;99(7):1042. <https://doi.org/10.1094/PDIS-12-14-1358-PDN>.
- Wu QF, Ding SW, Zhang YJ, Zhu SF. Identification of viruses and viroids by next-generation sequencing and homology-dependent and homology-independent algorithms. *Annu Rev Phytopathol.* 2015;53:425–44. <https://doi.org/10.1146/annurev-phyto-080614-120030>.
- Wylie SJ, Adams M, Chalam C, Kreuze J, López-Moya JJ, Ohshima K, et al. Erratum: ICTV virus taxonomy profile: Potyviridae. *J Gen Virol.* 2017;98(3):352–4. <https://doi.org/10.1099/jgv.0.000740>.
- Yamasaki S, Sakai J, Fuji S, Kamisoyama S, Emoto K, Ohshima K, et al. Comparisons among isolates of sweet potato featherly mottle virus using complete genomic RNA sequences. *Arch Virol.* 2010;155:795–800. <https://doi.org/10.1007/s00705-010-0633-0>.
- Yang J, Moeinzadeh MH, Kuhl H, Helmuth J, Xiao P, Haas S, et al. Haplotype-resolved sweet potato genome traces back its hexaploidization history. *Nat Plants.* 2017;3:696–703. <https://doi.org/10.1038/s41477-017-0002-z>.
- Zerbini FM, Briddon RW, Idris A, Martin DP, Moriones E, Navas-Castillo J, et al. ICTV virus taxonomy profiles: *Geminiviridae*. *J Gen Virol.* 2017;98(2):131–3. <https://doi.org/10.1099/jgv.0.000738>.
- Zhang S, Huang A, Zhou X, Li Z, Dietzgen RG, Zhou C, et al. Natural defect of a plant rhabdovirus glycoprotein gene: a case study of virus-plant co-evolution. *Phytopathology.* 2021;111(1):227–36. <https://doi.org/10.1094/PHTO-05-20-0191-FL>.

Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Ready to submit your research? Choose BMC and benefit from:

- fast, convenient online submission
- thorough peer review by experienced researchers in your field
- rapid publication on acceptance
- support for research data, including large and complex data types
- gold Open Access which fosters wider collaboration and increased citations
- maximum visibility for your research: over 100M website views per year

At BMC, research is always in progress.

Learn more biomedcentral.com/submissions

